

ICBBE 2020

NOVEMBER 06-09, 2020

KYOTO, JAPAN



PROCEEDINGS OF

2020 7th International Conference
on Biomedical and Bioinformatics Engineering

2020 7th International Conference on Biomedical and Bioinformatics Engineering (ICBBE 2020)

Table of Contents

Preface.....	vi
Conference Committee	vii

• ***Session 1 – Computational Biology and Biomedical Signal Analysis***

Dynamic Functional Connectivity and Graph Convolution Network for Alzheimer’s Disease Classification	1
<i>Xingwei An, Yutao Zhou, Yang Di and Dong Ming</i>	
Effective Evaluation of Clustering Algorithms on Single-Cell CNA data	5
<i>Marilisa M.M. Montemurro, Gianvito G.U. Urgese, Elena Grassi E.G Grassi, Carmelo Gabriele C.G.P., Andrea A.B. Bertotti and Elisa E.F. Ficarra</i>	
Expression and Methylation of Tumor Suppressor Gene DKK3 in Nasopharyngeal Carcinoma: A Datamining Study	12
<i>Zheng Xinyuan and Cen Dongzhi</i>	
Towards Data-Driven Modelling of Sumoylation Following Heat Shock	16
<i>Manyu Zhang, Yifei Zhang, Alice Zhao, Chun Guo and Lingzhong Guo</i>	
Using Hidden Markov Model for Identification Based on EEG Signals	22
<i>Wenxiao Zhong, Xingwei An, Yang Di, Lixin Zhang and Dong Ming</i>	
The Study of Voice Pathology Detection based on MFCC and SVM	27
<i>Yipeng Niu, Jiaming Cao, Fei Shen and Pengling Ren</i>	

• ***Session 2 – Medical Imaging and Image Diagnostics***

Improved Robustness in Water-Fat Separation in MRI using Conditional Adversarial Networks	31
<i>Chenfei Shen, Huajun She and Yiping Du</i>	
4th Order Tensors for Multi-fiber Resolution and Segmentation in White Matter	36
<i>Temesgen Bihonegn, Avinash Bansal, Jan Slovák and Sumit kaushik</i>	
Orientation and Distance Dependence of Pairwise Correlation in Macaque V1	43
<i>Lisha Hu, Qiyi Hu and Yao Chen</i>	
The Impact of Two Scatter Correction Methods on I-131 AC-SPECT Images using an Anthropomorphic Phantom with Variable Sizes of Thyroid Remnants	51
<i>Anastasia Hadjiconstanti, Konstantinos Michael, Theodoros Leontiou, Antonios Lontos, Savvas Frangos, George Demosthenous, Maria Lyra and Yiannis Parpottas</i>	

Optimization of Acoustic Noise for Single-Shot Echo-Planar Imaging by Varying Echo Spacing	59
<i>Zhenliang Lin, Qikang Li, Rui Wang, Guobin Li and Jie Luo</i>	
• <i>Session 3 – Pharmacy and Clinical Medicine</i>	
The Heart Failure Treatment of β -Blockers	66
<i>Su Jiujiu</i>	
A Review for Heart Regeneration	71
<i>Flora Yong Yu Chen</i>	
BIN1 Isoform 1 has Less Function in Promoting the Stability of TAT System in Adult Rat Cardiomyocytes	76
<i>Hao-Lin Zheng</i>	
Role of Anti-Müllerian Hormone (AMH) in Regulating Hypothalamus-Pituitary Function	82
<i>Yan Bin</i>	
Overview of Alzheimer’s Disease	87
<i>Wenlu Mao</i>	
Gait Stability Analysis with a Two-dimensional Dynamic Parameter	92
<i>Xing GAO, Fei Shen, Li Wang, Yingnan Ma, Haijun Niu and Yubo Fan</i>	
Modulation of Repetitive Transcranial Magnetic Stimulation on Mood and Cognitive Function in Simulated Weightlessness Rats	95
<i>Ling Wang, Jiajia Yang, Xi Xiao, Chenguang Zheng and Dong Ming</i>	
The Effect of High Fat Diets on Organic Acids in Mice	100
<i>Siyang Li</i>	
• <i>Session 4 – Molecular Biology and Biochemistry</i>	
The Molecular Dynamics Study on the Stability of Elk Prion Protein	104
<i>Ye Wang</i>	
The Molecular Dynamics Study on the PATHOGENICITY of Cystatin C Mutant	109
<i>Luying Pan</i>	
Biosensor-based Rapid Detection for Harmful Foodborne Pathogens	113
<i>Peitong Xu</i>	
Recent Insights into Proteomics in Plant Pathology	119
<i>Shiyu Lin</i>	
A Molecular View of Coronavirus Disease-2019 (COVID-19)	125
<i>Chenkai Jiang</i>	
Development and Application of CRISPR-Mediated Genetic Screening in Oncology	130
<i>Wanji Li</i>	

• **Session 5 – Cell Biology and Immunology**

Secretome and Ramiprilat Effects on Endothelial Progenitor Cells Proliferation in Chronic Coronary Syndrome Patient	136
<i>Yudi H. Oktaviono, Ferry Sandra, Achmad Lefi and Christian P. Budianto</i>	
Factors Affecting the Proliferation Ability of Cardiomyocytes	140
<i>JiaLiang Tan</i>	
The Viability Differences on Oocyte Vitrification Surrounded by Cumulus Cells and Oocyte Vitrification not Surrounded by Cumulus Cells after <i>In Vitro</i> Maturation	145
<i>AA Muhammad Nur Kasman, Budi Santoso, Widjiati Widjiati, Aucky Hinting, Ni Wajan Tirthaningsih, Reny I'tishom, Budi Utomo and Mochammad Sasmito Djati</i>	
A Novel Linear B-cell Epitope Prediction Method based on Position Entropy of Amino Acids	149
<i>Hong-guang Yang, Bin Cheng and Ling-Yun Liu</i>	
Ramiprilat Effects on Endothelial Progenitor Cells Migration is Increased by Human Umbilical Cord Blood-Mesenchymal Stem Cells derived Secretome	154
<i>Yudi Her Oktaviono, Ilma Alfia Isaridha, Ferry Sandra, Achmad Lefi and Agus Subagjo</i>	
The Research of Autophagy and Anti-Aging	160
<i>Baoshu An</i>	
Differential Response to the High Doses of Dimethyl Sulfoxide of the Several Human Cancer Cell Lines Cultured in 2D Monolayer, Decellularized Matrix, and 3D Spheroid cell Culture Systems	165
<i>Ekaterina YU Skorova, Eugenia Y Shabalina, Daria A Chudakova, Vladimir B Anikin, Igor V Reshetov, Ospan A Mynbaev and Elena V Petersen</i>	

• **Session 6 – Cancer Therapy and COVID-19**

Overview of Tumor Immunotherapy based on Indoleamine 2,3 Dioxygenase Inhibitors	171
<i>Xiangyu Hao</i>	
The Whole View of Therapies for Breast Cancer	176
<i>Cui Jie</i>	
Overview of Cancer Immunotherapy	180
<i>Zihan Wang</i>	
Study on Traditional Chinese Medicine for Treating COVID-19	184
<i>Jianqiu Wang</i>	
Viewing the Development Process of Coronavirus from the Diagnosis and Treatment of New Coronavirus	189
<i>Sarah Wan</i>	
View from Public Health to Molecular Biology on Coronavirus Disease 2019 (COVID-19)	194
<i>Yuru Li</i>	

Conference Committee

General Chairs

Prof. Kiyoshi Hoshino, University of Tsukuba, Japan

Assoc. Prof. Kuo-Yuan Hwa, National Taipei University of Technology, Taiwan

Program Chairs

Prof. Jose Nacher, Toho University, Japan

Prof. Qingli Li, East China Normal University, China

Technical Committees

Prof. Nagendra Kumar Kaushik, Kwangwoon University, South Korea

Prof. DoHoon Lee, Pusan National University, South Korea

Prof. Edwin Wang, National Research Council Canada/McGill University, Canada

Prof. Jun F. (James) Liang, Stevens Institute of Technology, USA

Prof. Trees-Juen Chuang, Genomics Research Center, Academia Sinica, Taiwan

Prof. Congo Tak Shing Ching, National Chung Hsing University, Taiwan

Prof. Muhammad Nawaz Iqbal, Pakistan Engineering Council, Pakistan

Assoc. Prof. Muchtaridi, Universitas Padjadjaran, Indonesia

Prof. Manoj R. Tarambale, Marathwada Mitra Mandal's College of Engineering, India

Dr. Binh P. Nguyen, Institute of High Performance Computing, A*STAR, Singapore

Prof. Alexander Polyakov, Sevastopol State University, Russia

Prof. Huaiqiu Zhu, Peking University, China

Assist. Prof. Jian-Guo Bau, Hungkuang University, Taiwan

Dr. Muhammad Arshad Malik, International Islamic University, Pakistan

Prof. Ekambaram Rajasekaran, Anna University, India

Prof. Ajitkumar Gorakhanath Patil, S.B.M.Polytechnic, India

Assoc. Prof. P. Shanmughavel, Bharathiar University Coimbatore, India

Prof. Direk Sueseenak, Srinakharinwirot University, Thailand

Dr. Mohd A. H. B. M. Adib, Universiti Malaysia Pahang, Malaysia

Prof. Shyam Narayan Labh, Nepal Armed Police Force School, Nepal

Prof. Satyabrata Aich, KIIT University, India

Assoc. Prof. Jin Zhang, Hunan Normal University, China

Assist. Prof. Nirmala Devi, National Institute of Technology Nagaland, India

Dr. Nung Kion Lee, Universiti Malaysia Sarawak, Malaysia

Assist. Prof. Steven Lim, Universiti Tunku Abdul Rahman, Malaysia

Assist. Prof. Yechun Ruan, The Hong Kong Polytechnic University, Hong Kong

Prof. Jya-Wei Cheng, National Tsing Hua University, Taiwan

Assoc. Prof. Shinya Nozaki, University of the Ryukyus, Japan

Assist. Prof. Li-Hui Lee, National Taipei University of Nursing and Health Sciences, Taiwan

Prof. Chiharu Ishii, Hosei University, Japan

Assist. Prof. Huang-Cheng Kuo, National Chiayi University, Taiwan

Assoc. Prof. Peng Du, Hangzhou Dianzi University, China

Dr. Irwansyah Idram, National Central University, Taiwan

Assist. Prof. Jhinuk Chatterjee, PES University, India

Assoc. Prof. Liudmila Davydova, Far Eastern Federal University, Russia

Assist. Prof. Napamane Kornthong, Thammasat University, Thailand

Assoc. Prof. Junichi Hoshino, University of Tsukuba, Japan

Prof. Yi-Horng Lai, Oriental Institute of Technology, Taiwan

Assoc. Prof. Khalid Malik, Oakland University, USA

Assist. Prof. Marco Frasca, Universita Degli Studi di Milano, Italy
Assoc. Prof. Huajun She, Shanghai Jiao Tong University, China
Assoc. Prof. Pufeng Du, Tianjin University, China
Assist. Prof. Sahar Al Seesi, Smith College, USA
Assist. Prof. Suparerk Janjarasjitt, Ubon Ratchathani University, Thailand
Prof. Boo Ho Voon, Universiti Teknologi MARA (UiTM), Malaysia
Assoc. Prof. Toru Hyakutake, Yokohama National University, Japan
Dr. Zhiping Liu, Anhui University of Technology, China
Dr. Duangdao Wichadakul, Chulalongkorn University, Thailand
Prof. Voon Boo Ho, Universiti Teknologi MARA (UiTM), Malaysia
Assoc. Prof. Hyakutake Toru, Yokohama National University, Japan
Prof. Aleksandr Poliakov, Sevastopol State University, Russia
Assoc. Prof. Jiajia Yang, Tianjin University, China
Lecturer Parichart Naruphontjirakul, King Mongkut's University of Technology Thonburi, Thailand
Assist. Prof. Huajun She, Shanghai Jiao Tong University, China
Assoc. Prof. Jie Luo, Shanghai Jiao Tong University, China
Assist. Prof. Wanwipa Siriwatwechakul, Thammasat University Pathum Thani, Thailand
Dr. Hongbin Li, Xianyang Vocational and Technical College, China
Prof. Yao Chen, Shanghai Jiao Tong University, China

Ramiprilat Effects on Endothelial Progenitor Cells Migration is Increased by Human Umbilical Cord Blood-Mesenchymal Stem Cells derived Secretome

Yudi Her Oktaviono
Department of Cardiology and
Vascular Medicine, Faculty of
Medicine, Universitas Airlangga, Prof
Moestopo Street 6-8, Surabaya,
Indonesia
yoktaviono@gmail.com

Ilma Alfia Isaridha
Department of Cardiology and
Vascular Medicine, Faculty of
Medicine, Universitas Airlangga, Prof
Moestopo Street 6-8, Surabaya,
Indonesia
ilmaalfia@gmail.com

Ferry Sandra
Department of Cardiology and
Vascular Medicine, Faculty of
Medicine, Universitas Airlangga, Prof
Moestopo Street 6-8, Surabaya,
Indonesia
ferrysandra@gmail.com

Achmad Lefi
Department of Cardiology and
Vascular Medicine, Faculty of
Medicine, Universitas Airlangga, Prof
Moestopo Street 6-8, Surabaya,
Indonesia
achmadlefi@gmail.com

Agus Subagjo
Department of Cardiology and
Vascular Medicine, Faculty of
Medicine, Universitas Airlangga, Prof
Moestopo Street 6-8, Surabaya,
Indonesia
agus.subagjo@fk.unair.ac.id

ABSTRACT

Endothelial progenitor cells (EPCs) have a critical role in angiogenesis and vasculogenesis of coronary artery disease (CAD) patients. Secretome of human Umbilical Cord Blood-Mesenchymal Stem Cell (hUCB-MSCs) can promote neovascularization. Ramiprilat is an active metabolite of ramipril that has shown benefit in cardiovascular disease. The effect of hUCB-MSCs-derived secretome alone or combination with ramiprilat on EPCs migration is not yet elucidated. This study aimed to identify the effect of hUCB-MSC derived secretome and its combination with ramiprilat on EPCs migration. EPCs were collected from peripheral blood of CAD patient and cultured in the Stemline II medium. Cultured EPCs were then divided into groups of control, ramiprilat 10 μ mol, hUCB-MSCs derived secretome (2%, 10%, and 20%), and its combination. The migration of EPCs was assessed using a Boyden chamber assay. Ramiprilat and hUCB-MSCs-derived secretome at all doses increase EPCs migration in dose-dependent manner. Combination of hUCB-MSCs-derived secretome at dose 10% and 20% and ramiprilat significantly increase migrated cells compared to ramiprilat only and secretome only group ($p < 0.001$). In conclusion, hUCB-MSCs-derived secretome and ramiprilat enhance EPCs migration and combination of those two substances furtherly increased the migrated cells. hUCB-MSCs-derived secretome has the potential as regenerative treatment for CAD patients.

Permission to make digital or hard copies of all or part of this work for personal or classroom use is granted without fee provided that copies are not made or distributed for profit or commercial advantage and that copies bear this notice and the full citation on the first page. Copyrights for components of this work owned by others than ACM must be honored. Abstracting with credit is permitted. To copy otherwise, or republish, to post on servers or to redistribute to lists, requires prior specific permission and/or a fee. Request permissions from permissions@acm.org.

ICBBE '20, November 06–09, 2020, Kyoto, Japan

© 2020 Association for Computing Machinery.

ACM ISBN 978-1-4503-8822-1/20/11...\$15.00

<https://doi.org/10.1145/3444884.3444914>

CCS CONCEPTS

• **Applied computing**; • **Life and medical sciences**; • **System biology**;

KEYWORDS

endothelial progenitor cells, human umbilical cord blood, mesenchymal stem cells, secretome, migration, ramiprilat

ACM Reference Format:

Yudi Her Oktaviono, Ilma Alfia Isaridha, Ferry Sandra, Achmad Lefi, and Agus Subagjo. 2020. Ramiprilat Effects on Endothelial Progenitor Cells Migration is Increased by Human Umbilical Cord Blood-Mesenchymal Stem Cells derived Secretome. In *2020 7th International Conference on Biomedical and Bioinformatics Engineering (ICBBE '20)*, November 06–09, 2020, Kyoto, Japan. ACM, New York, NY, USA, 6 pages. <https://doi.org/10.1145/3444884.3444914>

1 INTRODUCTION

Coronary artery disease (CAD) has a high number of morbidity and mortality around the world. Coronary events are expected to occur in more than one million individuals in United States [1]. While CAD therapies are increasingly progressing in terms of pharmacological and percutaneous intervention techniques, there are some patients that do not take benefits of this advancement therapy including patients with refractory angina [15, 25]. These patients need therapy modalities which can improve their quality of life.

Endothelial progenitor cells (EPCs) are multipotent cell that has an important role in the pathophysiology of coronary artery disease. EPCs are able to differentiate into mature endothelial cell, contributes to reendothelialization after endothelial injury and therefore improve endothelial function. EPCs also play critical role in angiogenesis and vasculogenesis. To do this role, EPCs need to mobilize from bone marrow to circulating blood and migrate to

injured site [20, 31]. Unfortunately, people with low EPCs level, including CAD patients, have an impairment in forming collateral and compensates for the presence of stenosis [5]. Thereby, improving circulating EPCs number and function provide therapeutic option for CAD patients.

Human umbilical cord blood mesenchymal stem cells (hUCB-MSCs) secrete molecules to extracellular space named secretome, that contain soluble proteins, exosomes and micro vesicles [6, 9, 24]. This secretome promotes neovascularization, angiogenesis [6, 32], and improves cardiac systolic function [34]. hUCB-MSCs as source of secretome has advantage in high proliferation capacity and less invasive collecting method [14]. Another researches have shown the migration enhancing capacity of using another source of secretome [3, 12] but the effect of hUCB-MSCs derived secretome on EPCs migration has not yet established. Ramipril, with active metabolite ramiprilat, is an angiotensin converting enzyme inhibitor (ACE-I) that has been shown to increase EPCs proliferation and migration [21, 22]. This study is aimed to identify the effect of hUCB-MSCs derived secretome and its combination with ramiprilat on EPC migration.

2 METHODS

2.1 Sample criteria

This experiment used blood samples that were obtained from chronic coronary syndrome patient at dr. Soetomo General Hospital Surabaya Outpatient Clinic. The inclusion criteria for this study were male, aged 40–59 years old, had conducted coronary angiography that showed >50% stenosis of left main coronary artery or in the other coronary arteries showed 70% stenosis. We exclude patients with acute coronary syndrome, acute limb ischemia, diabetes mellitus and anemia. This study protocol had an ethical clearance from the Health Research Ethics Committee of Dr. Soetomo General Hospital Surabaya. The subject had signed informed consent before recruited. All details which included personal information were omitted.

2.2 Secretome preparation

The preparation of hUCB-MSCs derived secretome was done in accordance to previous study [26]. hUCB-MSCs cell line (3H Biomedical AB, Uppsala, Sweden) was cultured in Mesencult media (Stem-Cell Technologies Inc., Vancouver, Canada) which contained penicillin and streptomycin. While the confluency was reaching 80%, the media was replaced with the newer media with no supplementation. hUCB-MSCs with supplement-free media then incubated for 24 hours. After incubation, the media was collected and centrifuged. Supernatant that was resulted from centrifugation was used as a conditioned medium that contained secretome.

2.3 EPCs isolation and culture

EPCs were collected from mononuclear cells (MNCs) of the peripheral blood of CAD patient. Forty milliliters of blood were diluted with phosphate buffer saline with 2% fetal bovine serum then ficoll histopaque was added. Centrifugation of the mixture was done until peripheral blood MNCs (PBMNCs) layer was formed. PBMNCs were cultured with basal Stemline II hematopoietic stem cell

expansion medium (Sigma-Aldrich, St. Louis, MO, USA) supplemented with 15% fetal bovine serum and several growth factors in the fibronectin-coated 6-well plates. The culture was maintained at 37°C with 5% CO₂ in a humidified atmosphere. Non-adherent cells were removed, and fresh medium was added.

2.4 EPCs identification

After three days of isolation and culture, EPCs were confirmed using immunofluorescence microscope examination with FITC-labeled anti-human CD34 antibody (Biolegend, USA) staining.

2.5 EPCs treatment

Cultured EPCs were divided into control group, treatment with 10 μ mol Ramiprilat, various concentrations of hUCB-MSCs-derived secretome (2%, 10%, and 20%) and combination of 10 μ mol Ramiprilat and each dose of secretome. The control group was not treated with secretome or ramiprilat. For the Ramiprilat group, cell culture was incubated for 48 hours before transferring to the transwell.

2.6 Migration assay

This experiment used Costar® Transwell® Permeable Support (Corning, USA) with a 3.0 μ m pore size membrane. EPCs migration was assessed using a Boyden chamber assay method. A total of 5×10^5 EPCs were placed at the upper chamber with basal media and the lower chamber was supplemented with basal media and secretome. The culture was incubated at 37°C for 24 hours. The non-migratory cells were removed manually. The migratory EPCs below the upper chamber were fixed with 3.7% paraformaldehyde and permeabilized with methanol. Migrated EPCs were stained with Giemsa staining and calculated.

2.7 Statistical analysis

Data analyses were done using SPSS Statistics 23.0 from IBM to detect significance level at $p < 0.05$. Data distribution was evaluated using Kolmogorov Smirnov test and comparison between groups were calculated using one-way ANOVA test. Correlation between variables was obtained using Spearman correlation followed by a linear regression test.

3 RESULTS

3.1 EPCs identification

Identification of EPCs was done using immunofluorescence and light microscope. Positive CD34 expression was used to mark EPCs. Under a light microscope, EPCs were demonstrated as spindle-shaped cells (Figure 1).

3.2 EPCs migration

This experiment showed that ramiprilat and all doses of hUCB-MSCs derived secretome significantly increased EPCs migration compared to control group ($p < 0.001$). hUCB-MSCs derived secretome increase EPCs migration in dose dependent manner (Figure 2). Ramiprilat 10 μ mol had significantly higher EPCs migration than secretome 2% (33.80 ± 2.49 vs 17.20 ± 1.92 , $p < 0.001$) but no statistically significant difference was observed between ramiprilat and secretome 10% (33.80 ± 2.49 vs 27.00 ± 4.00 , $p > 0.05$). However,

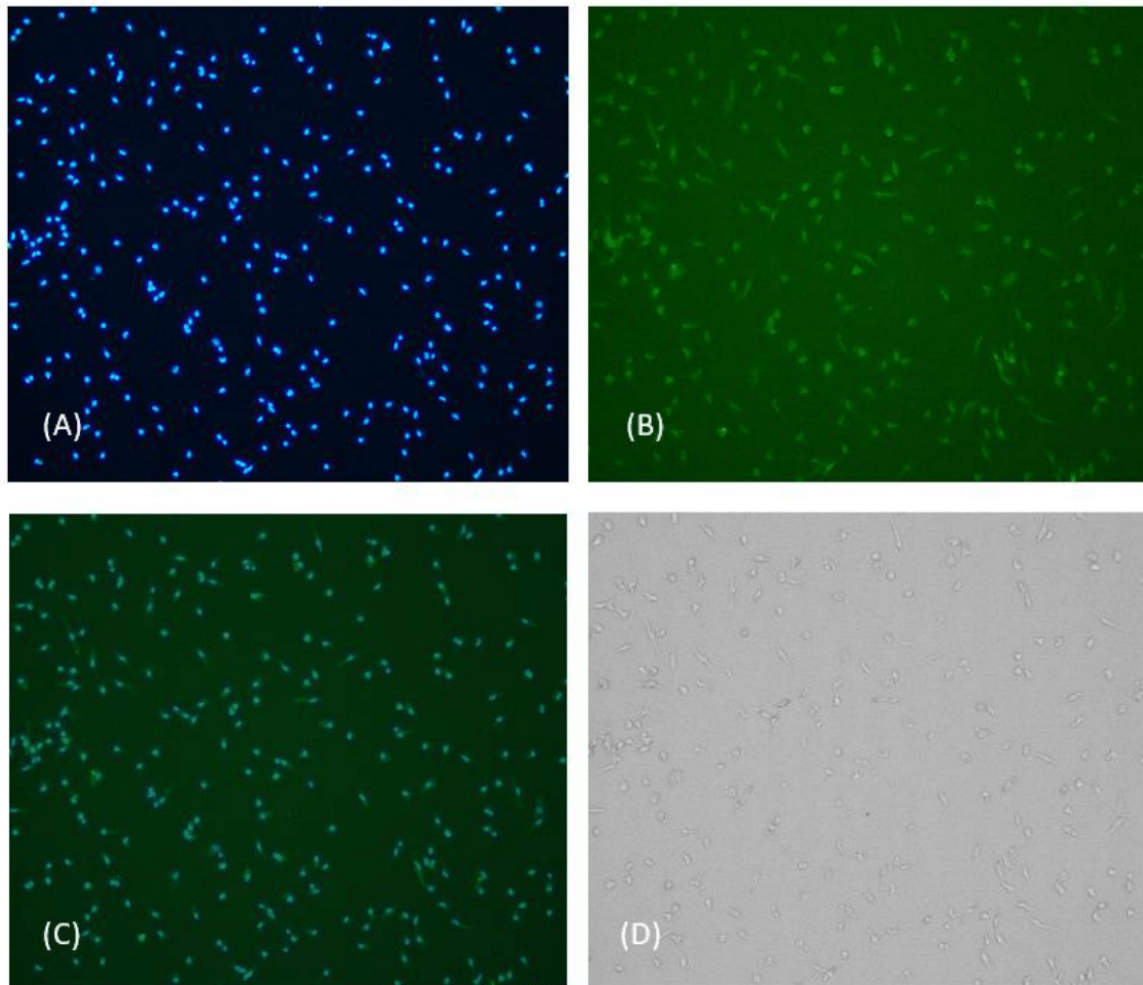


Figure 1: Microscopic picture of endothelial progenitor cells (EPCs) after three days incubation and culture (200x magnification) (A) Staining with diamidino-2-phenylindole (DAPI) showing nuclear image of EPCs with blue fluorescence (B) FITC-labelled anti-CD34 antibody staining of EPCs (C) Merged view of A and B (D) Spindle shape of EPCs were shown in light microscope

secretome 20% showed a significantly higher migration compared to the ramiprilat group (51.00 ± 5.15 vs 33.80 ± 2.49 , $p < 0.001$).

Spearman correlation showed a significant and strong correlation between hUCB-MSCs-derived secretome treatment with EPCs migration ($r = 0.946$; $p < 0.001$). The linear regression test showed R-square of 0.877. This indicated that hUCB-MSCs-derived secretome treatment was responsible for 87.7% increase in EPCs migration.

Combination of ramiprilat $10 \mu\text{mol}$ and hUCB-MSCs derived secretome at all doses showed significant enhancement of EPCs migration compared to secretome only group (30.00 ± 4.06 vs 17.20 ± 1.92 , 55.00 ± 4.42 vs 27.00 ± 4.00 , 69.00 ± 7.65 vs 51.00 ± 5.15 , $p < 0.001$). Ramiprilat only group showed higher number of EPCs migration compared to combination of Ramiprilat and secretome 2% but not statistically significant (33.80 ± 2.49 vs 30.00 ± 4.06 , $p > 0.05$). Combination of ramiprilat and secretome at 10% and 20% concentration were significantly superior to ramiprilat only group (55.00 ± 4.42

vs 33.80 ± 2.49 and 69.00 ± 7.65 vs 33.80 ± 2.49 , $p < 0.001$). The combination hUCB-MSCs derived secretome 20% and ramiprilat had the highest number of migrated EPCs compared to another groups (Figure 3).

4 DISCUSSION

The result of present study demonstrated that hUCB-MSCs derived secretome, ramiprilat, and combination of both of them enhance EPCs migration. Secretome increased EPCs migration in dose dependent manner. Combination of ramiprilat and high dose secretome has the highest number of EPCs migration compared to ramiprilat alone or secretome alone. This synergistic effect might be beneficial for treatment in chronic coronary syndrome patients.

Mesenchymal stem cells secretome contained pro angiogenic factors including insulin-like growth factor (IGF), interleukin-6 (IL-6), stromal cell-derived factor-1 (SDF-1), prostaglandin E2 (PGE2),

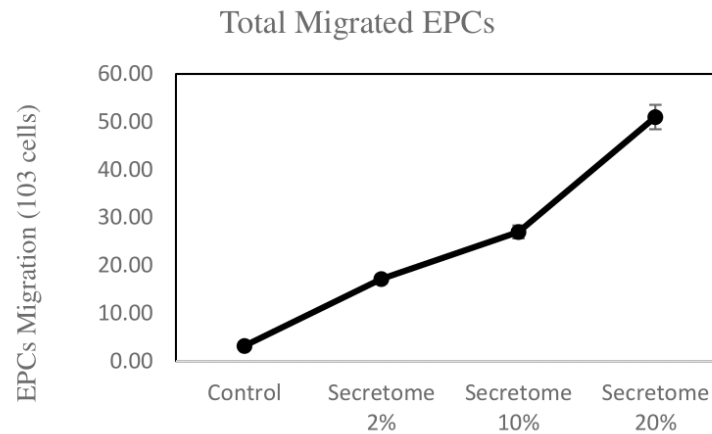


Figure 2: Human umbilical cord blood mesenchymal stem cells (hUCB-MSCs) derived secretome improves endothelial progenitor cells (EPCs) migration in dose dependent manner. EPCs were treated with hUCB-MSCs derived secretome at doses 2%, 10% and 20%. EPCs migration was counted using Boyden chamber assay.

vascular endothelial growth factor (VEGF), vascular cell adhesion protein 1 (VCAM-1), microvesicles and exosomes [6, 7, 9, 24]. Secretome played role in cardiac tissue preservation, formation of new vessels in damaged tissue, immunomodulation and cardiac regeneration [6, 32, 33]. The results of this research are in line with previous study using placental-derived MSCs (PL-MSCs). PL-MSCs soluble factors significantly enhance EPC migration. Several secreted proteins identified as candidates for EPC migration enhancing factor [12, 13]. Another study using human amniotic membrane-derived mesenchymal stromal cells (hAMCs) secretome treatment also show increased EPCs migration in a dose-dependent manner [3]. While this research shows that hUCB-MSCs derived secretome does increase EPCs migration, the exact mechanism has not yet been established.

During ischemic condition, the affected area releases variety of signal factor including proangiogenic chemoattractant which triggers homing of EPCs to the ischemic area and enhance angiogenesis and vasculogenesis [20, 31]. Elevation of EPCs was seen in myocardial infarction and this increase is in line with increase of VEGF. VEGF increased EPCs migration by binding to VEGFR1 and VEGFR2, shifting G protein signaling toward RAC and RHO which was important for cytoskeletal rearrangement [8, 18, 29].

SDF-1 was the most potent chemoattractant for EPCs. SDF-1 concentration gradient from peripheral blood to ischemic area played a critical role in EPCs migration [19]. SDF-1 bound to the CXCR4 and activated Rac GTPase protein. Its downstream pathway regulates cellular polarity and cytoskeleton changes that accomplish directional migration [27]. The important role of CXCR4/SDF-1 axis was regulated by hypoxia-inducible factors 1 α (HIF-1 α) [30]. Another research suggested that SDF-1-induced EPCs migration was mediated through the PI3K/Akt/eNOS signal transduction pathway [35]. hUCB-MSCs derived secretome contained SDF-1 which can increase chemoattractant gradient. Giving hUCB-MSCs derived secretome with a higher concentration will increase the gradient so that EPCs will move faster toward the ischemic area. This is

indicated in our result which EPCs migration is increase along with the increase in secretome doses.

This study shows that ramiprilat treatment increased EPCs migration better than hUCB-MSC derived secretome at dose 2% and 10%. Ramiprilat is an active metabolite of ramipril, a non-sulfhydryl angiotensin converting enzyme (ACE) inhibitor that blocks the conversion of angiotensin I to angiotensin II, and inhibit degradation of bradykinin [2]. Previous studies had shown that increased in EPCs migration occurred after 7 days of ramipril administration in stable coronary artery disease patients [21]. The underlying mechanism was thought to be related to the bradykinin pathway. ACE inhibitors block the degradation of B2R agonist and enhancing B2R signaling [4]. EPCs stimulation with bradykinin can increase the formation of filopodia and accelerate EPC migration [16]. Bradykinin has an important role in vascular function and involves in eNOS expression through activation of PI3K/Akt-dependent and independent pathway [11, 28].

The combination of ramiprilat with hUCB-MSCs derived secretome shows a synergistic effect where the amount of migrated EPCs increase significantly. The combination of ramiprilat and 20% hUCB-MSC derived secretome shows the highest EPCs migration number, exceeding ramiprilat only group and secretome only group. It is speculated that the combination of these two substances accelerate the migration of EPCs through various mechanism. Apart from the paths mentioned above, hUCB-MSCs also contains a high level of the exosome that has been investigated as proven to reduce myocardial ischemia by inducing neovascularization and increasing vascular tube formation [32]. hUCB-MSCs derived secretome also has anti-inflammatory and antioxidant properties that is expected to improve EPCs migration [10, 17]. Previous research showed that antioxidants can increase EPCs migration [23].

5 STUDY LIMITATIONS

This research has not yet identified which molecules contained in the hUCB-MSCs derived secretome that has major influence

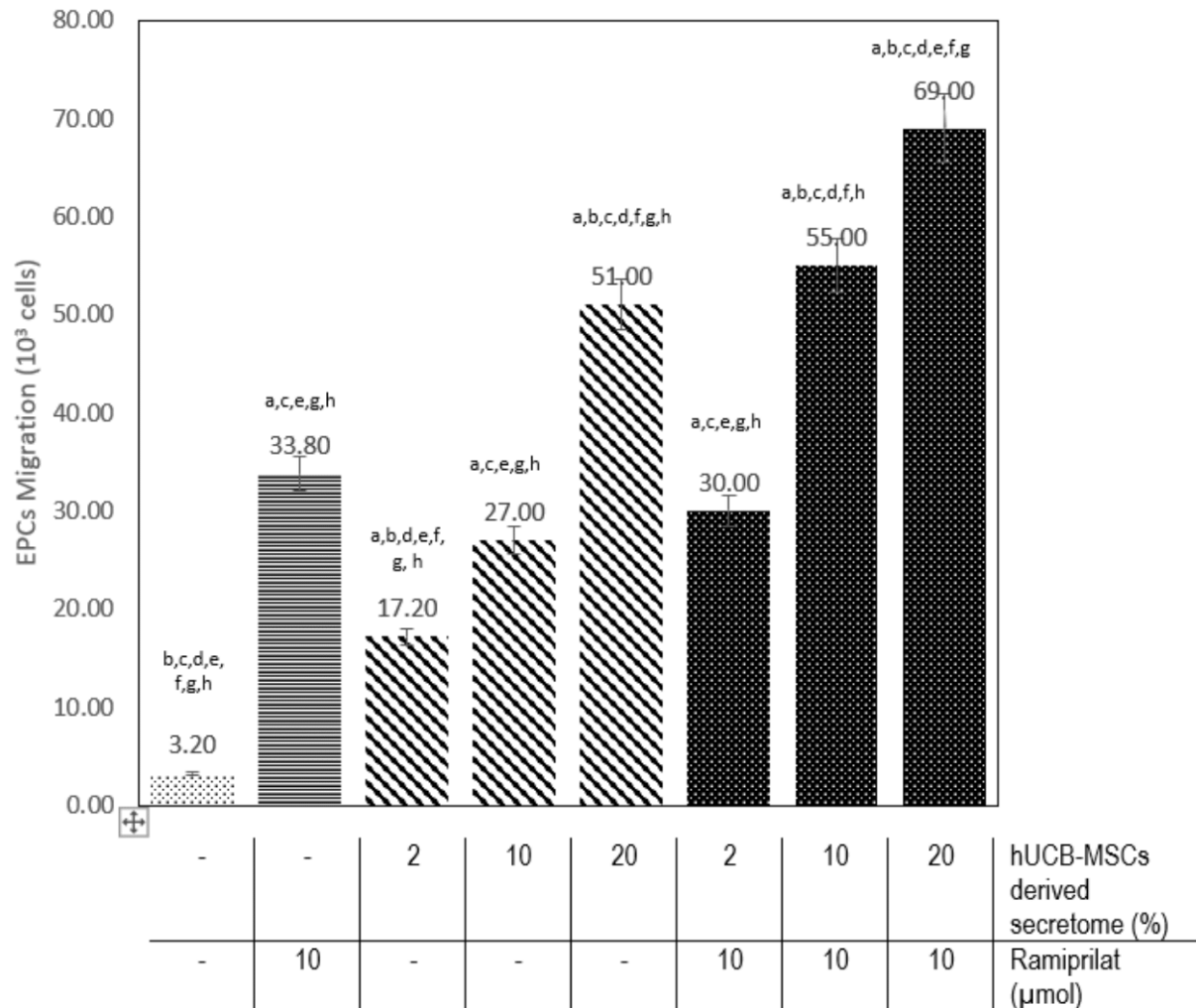


Figure 3: Comparison of EPCs migration between all groups. a: EPCs migration significantly increased compared to control group ($p<0.001$), **b:** EPCs migration significantly increased compared to the 10 µmol ramiprilat group ($p<0.001$), **c:** EPCs migration significantly increased compared to the 2% hUCB-MSCs derived secretome group, ($p<0.001$), **d:** EPCs migration significantly increased compared to the 10% hUCB-MSCs derived secretome group ($p<0.001$), **e:** EPCs migration significantly increased compared to the 20% hUCB-MSCs derived secretome group ($p<0.001$), **f:** EPCs migration significantly increased compared to the combination of 2% hUCB-MSCs derived secretome and ramiprilat group, ($p<0.001$), **g:** EPCs migration significantly increased compared to the combination of 10% hUCB-MSCs derived secretome and ramiprilat group, ($p<0.001$) and **h:** EPCs migration significantly increased compared to the combination of 20% hUCB-MSCs derived secretome and ramiprilat group, ($p<0.001$).

in EPCs migration. Further research is needed to verify various mechanisms speculated to improve EPCs migration in hUCB-MSCs derived secretome group and in combination group.

6 CONCLUSION

hUCB-MSCs-derived secretome and ramiprilat enhance EPCs migration. Combination of those two substances furtherly increased

the migrated cells. hUCB-MSCs-derived secretome has the potential as a cardiovascular regenerative treatment for the patient with CAD.

ACKNOWLEDGMENTS

This research was funded by Indonesian Ministry of Research and Technology – National Research and Innovation Foundation (Kemeterian Riset dan Teknologi – Badan Riset dan Inovasi Nasional)

REFERENCES

- [1] Emelia J. Benjamin, Paul Muntner, Alvaro Alonso, et al. 2019. Heart Disease and Stroke Statistics-2019 Update: A Report From the American Heart Association. *Circulation* 139, 10: e56–e528.
- [2] Siobhan Boyle, Gerry McKay, and Miles Fisher. 2013. Ramipril. 39–42.
- [3] Patrizia Danieli, Giuseppe Malpasso, Maria Chiara Ciuffreda, et al. 2015. Conditioned Medium From Human Amniotic Mesenchymal Stromal Cells Limits Infarct Size and Enhances Angiogenesis. *STEM CELLS Translational Medicine* 4, 5: 448–458.
- [4] Ervin G. Erdős, Fulong Tan, and Randal A. Skidgel. 2010. Angiotensin I-converting enzyme inhibitors are allosteric enhancers of kinin B1 and B2 receptor function. *Hypertension* 55, 2: 214–220.
- [5] Gian Paolo Fadini, Carlo Agostini, Saverio Sartore, and Angelo Avogaro. 2007. Endothelial progenitor cells in the natural history of atherosclerosis. *Atherosclerosis* 194, 1: 46–54.
- [6] Clara Gallina, Valentina Turinetto, and Claudia Giachino. 2015. A New Paradigm in Cardiac Regeneration: The Mesenchymal Stem Cell Secretome. *Stem Cells International* 2015.
- [7] Carl Harrell, Crissy Fellabaum, Nemanja Jovicic, Valentin Djonov, Nebojsa Arsenijevic, and Vladislav Volarevic. 2019. Molecular Mechanisms Responsible for Therapeutic Potential of Mesenchymal Stem Cell-Derived Secretome. *Cells* 8, 5: 467.
- [8] Brian R. Hoffmann, Jordan R. Wagner, Anthony R. Prisco, Agnieszka Janiak, and Andrew S. Greene. 2013. Vascular endothelial growth factor-A signaling in bone marrow-derived endothelial progenitor cells exposed to hypoxic stress. *Physiological Genomics* 45, 21: 1021–1034.
- [9] Hyo Jeong Hong, Hocon Kim, Chae Woon Park, et al. 2009. Cytokine Secretion Profiling of Human Mesenchymal Stem Cells by Antibody Array. *International Journal of Stem Cells* 2, 1: 59–68.
- [10] Hye Jin Jin, Yun Kyung Bae, Miyeon Kim, Soon-jae Kwon, and Hong Bae Jeon. 2013. Comparative Analysis of Human Mesenchymal Stem Cells from Bone Marrow, Adipose Tissue, and Umbilical Cord Blood as Sources of Cell Therapy. 1: 17986–18001.
- [11] Hong Ju, Virginia J. Venema, Mario B. Marrero, and Richard C. Venema. 1998. Inhibitory interactions of the bradykinin B2 receptor with endothelial nitric-oxide synthase. *Journal of Biological Chemistry* 273, 37: 24025–24029.
- [12] Witchayaporn Kamprom, Pakpoom Kheolamai, Yaowalak U-Pratya, et al. 2016. Endothelial Progenitor Cell Migration-Enhancing Factors in the Secretome of Placental-Derived Mesenchymal Stem Cells. *Stem Cells International* 2016.
- [13] Witchayaporn Kamprom, Pakpoom Kheolamai, Yaowalak U-Pratya, et al. 2016. Effects of mesenchymal stem cell-derived cytokines on the functional properties of endothelial progenitor cells. *European Journal of Cell Biology* 95, 3–5: 153–163.
- [14] Susanne Kern, Hermann Eichler, Johannes Stoeve, Harald Klüter, and Karen Bieback. 2006. Comparative Analysis of Mesenchymal Stem Cells from Bone Marrow, Umbilical Cord Blood, or Adipose Tissue. *Stem Cells* 24, 5: 1294–1301.
- [15] Juhani Knuuti, William Wijns, Stephan Achenbach, et al. 2020. 2019 ESC guidelines for the diagnosis and management of chronic coronary syndromes. *European Heart Journal* 41, 3: 407–477.
- [16] Nicolle Kränkel, Rajesh G. Katare, Mauro Siragusa, et al. 2008. Role of kinin b2 receptor signaling in the recruitment of circulating progenitor cells with neovascularization potential. *Circulation Research* 103, 11: 1335–1343.
- [17] Kuo Hua Lee, Wei Cheng Tseng, Chih Yu Yang, and Der Cherng Tarnq. 2019. The Anti-Inflammatory, Anti-Oxidative, and Anti-Apoptotic Benefits of Stem Cells in Acute Ischemic Kidney Injury. *International Journal of Molecular Sciences* 20.
- [18] Bin Li, Emerson E. Sharpe, Amanda B. Maupin, et al. 2006. VEGF and PlGF promote adult vasculogenesis by enhancing EPC recruitment and vessel formation at the site of tumor neovascularization. *The FASEB Journal* 20, 9: 1495–1497.
- [19] Da Wei Li, Zhi Qiang Liu, Jun Wei, Ying Liu, and Lin Sen Hu. 2012. Contribution of endothelial progenitor cells to neovascularization (review). *International Journal of Molecular Medicine* 30, 5: 1000–1006.
- [20] H. Masuda, A. Kawamoto, M. Ii, and T. Asahara. 2016. Endothelial Progenitor Cells for Vascular Medicine. *Regenerative Medicine for Peripheral Artery Disease*: 71–90.
- [21] Tao Qian Min, Chen Jun Zhu, Wang Xing Xiang, Zhu Jun Hui, and Shang Yun Peng. 2004. Improvement in endothelial progenitor cells from peripheral blood by ramipril therapy in patients with stable coronary artery disease. *Cardiovascular Drugs and Therapy* 18, 3: 203–209.
- [22] Patrick Müller, Andrey Kazakov, Philippe Jagoda, Alexander Semenov, Michael Böhm, and Ulrich Laufs. 2009. ACE inhibition promotes upregulation of endothelial progenitor cells and neoangiogenesis in cardiac pressure overload. *Cardiovascular Research* 83, 1: 106–114.
- [23] Yudi Her Oktaviono, Muhammad Rafdi Amadis, and Makhyhan Jibril Al-Farabi. 2020. High dose allicin with Vitamin C improves EPCs migration from the patient with coronary artery disease. *Pharmacognosy Journal* 12, 2: 232–235.
- [24] Jolene Phelps, Amir Sanati-Nezhad, Mark Ungrin, Neil A. Duncan, and Arindom Sen. 2018. Bioprocessing of mesenchymal stem cells and their derivatives: Toward cell-free therapeutics. *Stem Cells International* 2018, iii.
- [25] Paul A. Sainsbury, Michael Fisher, and Ranil De Silva. 2017. Alternative interventions for refractory angina. *Heart* 103, 23: 1911–1922.
- [26] Ferry Sandra, Janti Sudiono, Elina Ardiani Sidharta, et al. 2014. Conditioned Media of Human Umbilical Cord Blood Mesenchymal Stem Cell-derived Secretome Induced Apoptosis and Inhibited Growth of HeLa Cells. *The Indonesian Biomedical Journal* 6, 1: 57.
- [27] Li Shen, Yongxing Gao, Juying Qian, Aijun Sun, and Junbo Ge. 2011. A novel mechanism for endothelial progenitor cells homing: The SDF-1/CXCR4-Rac pathway may regulate endothelial progenitor cells homing through cellular polarization. *Medical Hypotheses* 76, 2: 256–258.
- [28] Jin Bo Su. 2018. Role of Bradykinin in the Regulation of Endothelial Nitric Oxide Synthase Expression by Cardiovascular Drugs. *Current Pharmaceutical Design* 23, 40: 6215–6222.
- [29] Paul E. Szmitko, Paul W.M. Fedak, Richard D. Weisel, Duncan J. Stewart, Michael J.B. Kutryk, and Subodh Verma. 2003. Endothelial progenitor cells: New hope for a broken heart. *Circulation* 107, 24: 3093–3100.
- [30] Tran Cam Tu, Masumi Nagano, Toshiharu Yamashita, et al. 2016. A Chemokine Receptor, CXCR4, Which Is Regulated by Hypoxia-Inducible Factor 2 α , Is Crucial for Functional Endothelial Progenitor Cells Migration to Ischemic Tissue and Wound Repair. *Stem Cells and Development* 25, 3: 266–276.
- [31] Carmen Urbich and Stefanie Dimmeler. 2004. Endothelial progenitor cells: Characterization and role in vascular biology. *Circulation Research* 95, 4: 343–353.
- [32] Francisco J. Vizoso, Noemi Eiro, Sandra Cid, Jose Schneider, and Roman Perez-Fernandez. 2017. Mesenchymal stem cell secretome: Toward cell-free therapeutic strategies in regenerative medicine. *International Journal of Molecular Sciences* 18, 9.
- [33] Mingjun Wu, Ruifan Zhang, Qing Zou, et al. 2018. Comparison of the Biological Characteristics of Mesenchymal Stem Cells Derived from the Human Placenta and Umbilical Cord. *Scientific Reports* 8, 1.
- [34] Yuanyuan Zhao, Xiaoxian Sun, Wenming Cao, et al. 2015. Exosomes Derived from Human Umbilical Cord Mesenchymal Stem Cells Relieve Acute Myocardial Ischemic Injury. *Stem Cells International* 2015.
- [35] Hao Zheng, Guosheng Fu, Tao Dai, and He Huang. 2007. Migration of endothelial progenitor cells mediated by stromal cell-derived factor-1 α /CXCR4 via PI3K/Akt/eNOS signal transduction pathway. *Journal of Cardiovascular Pharmacology* 50, 3: 274–280.